



Exogenous vs. endogenous γ -glutamyltransferase activity: Implications for the specific determination of S-nitrosoglutathione in biological samples

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ABSTRACT

The determination of S-nitrosoglutathione (GSNO) levels in biological fluids is controversial, partly due to the laborious sample handling and multiple pretreatment steps required by current techniques. GSNO decomposition can be effected by the enzyme gamma-glutamyltransferase (GGT), whose involvement in GSNO metabolism has been suggested. We have set up a novel analytical method for the selective determination and speciation of GSNO and its metabolite S-nitrosocysteinylglycine, based on liquid chromatography separation coupled to on-line enzymatic hydrolysis of GSNO by commercial GGT. In a post-column reaction coil, GGT allows the specific hydrolysis of the γ -glutamyl moiety of GSNO, and the S-nitrosocysteinylglycine (GCNO) thus formed is decomposed by copper ions originating oxidized cysteinylglycine and nitric oxide (NO). NO immediately reacts with 4,5-diaminofluorescein (DAF-2) forming a triazole derivative, which is detected fluorimetrically. The limit of quantitation (LOQc) for GSNO and GCNO in plasma ultrafiltrate was 5 nM, with a precision (CV) of 1–6% within the 5–1500 nM dynamic linear range.

The method was applied to evaluate the recovery of exogenous GSNO after addition of aliquots to human plasma samples presenting with different total GGT activities. By inhibiting GGT activity in a time dependent manner, it was thus observed that the recovery of GSNO is inversely correlated with plasmatic levels of endogenous GGT, which indicates the need for adequate inhibition of endogenous GGT activity for the reliable determination of endogenous GSNO.

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Introduction

It is widely accepted that S-nitrosothiols (RSNOs)¹ are important intermediates in nitric oxide (NO) metabolism, though their significance in pathophysiological phenomena still remains under debate. Several methods have been published in recent years, all including rather aggressive and non-specific decomposition procedures [photolysis, HgCl₂, HgCl₂/V(III), KI/I₂, Cys/KI/Cu(I) etc., liable to cause undesired interactions with components of biological matrices[1]. Besides, a major obstacle in assessing reference values

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¹ RSNOs, S-nitrosothiols; GSNO, S-nitrosoglutathione; GGT, gamma-glutamyltransferase; GCNO, S-nitrosocysteinylglycine; LOQc, limit of quantitation; NO, nitric oxide; GG, glycylglycine; LC, liquid chromatography; DAF-2, diaminofluorescein; DMSO, dimethyl sulfoxide; NEM, N-ethylmaleimide; DTPA, diethylenetriamine-pentaacetic acid pentasodium salt; SBC, serine/borate complex; DAD, diode array detector; FD, fluorimetric detector.

of RSNOs in complex biological matrices is represented by the lack of consolidated procedures for sampling/storage to preserve RSNO stability in the pre-analytical step, as well as of validated analytical methods/protocols and certified reference material [1]. As a consequence, the range of basal RSNO levels reported in various studies has covered more than three orders of magnitude, between low nanomolar (<1–62 nM) and low-micromolar/micromolar levels [1–3].

GGT is a central enzyme for the synthesis and degradation of GSH [4]. GGT can also cause decomposition of GSNO [5,6], and the metabolite GCNO is more susceptible than GSNO to copper ion-catalysed decomposition [7]. Thus, in the presence of glycylglycine (GG) as acceptor, the following reactions occur *in vitro*:



It was reported that the reaction of Cu(II) with GCNO is fast, with half-lives of about 5 s, while the reaction with GSNO is 3–4 orders of magnitude slower [7]. By *in vitro* kinetics analysis we have recently shown that bovine kidney GGT has a K_m value for GSNO of 0.398 ± 0.031 mM, comparable with the K_m value for GSH (0.4 mM) [8].

In the present study we have set up a new analytical method for the specific determination and speciation of GSNO and GCNO, based on liquid chromatography (LC) and an on-line, post-column reaction system. In the latter, conditions are adjusted for enzymatic decomposition of GSNO by commercial bovine kidney GGT (reaction (1)), fast decomposition of its product GCNO by copper ions (reaction (2)) [9], and reaction of NO thus formed with 4,5-diaminofluorescein (DAF-2) (reaction (3)) [10,11]:



The triazole derivative, DAF-2T, is detected and quantified fluorimetrically. The procedure allowed the separation of GSNO and GCNO from compounds present in complex matrices and in the presence of ascorbic acid and uric acid, which are known to interfere with NO detection [12–14]. Besides, data obtained with plasma samples confirmed that a loss of GSNO may occur during pre-analytical handling when endogenous GGT present is not inhibited.

Materials and methods

Chemicals

Analytical reagent-grade chemicals were used without further purification. GGT (G-8040, Type I crude from bovine kidney) and glycylglycine (GG, G-1002) were purchased from Sigma–Aldrich (Sigma, Chemical Co., Milan, Italy). Copper sulphate anhydrous (2791) was purchased from Merck (Laborchimica, Firenze, Italy). Five millimolar solution of 4,5-Diaminofluorescein (DAF-2) solution in dimethyl sulfoxide (DMSO) (251510) and its triazole derivative (DAF-2T, 251510, 0.5 mM in DMSO) were purchased from Calbiochem (La Jolla CA, USA).

N-ethylmaleimide (NEM, 04259 BioChemika Ultra, $\geq 99.0\%$), DL-serine (84980), boric acid (B-0252) and diethylenetriamine-pentaacetic acid pentasodium salt (DTPA, 17969) were purchased from Fluka (Fluka GmbH, Buchs, Switzerland). 50 mM stock solution of ethylenediaminetetraacetic acid (405501, EDTA disodium salt, RPE) was purchased from Carlo Erba (Milan, Italy). Phosphate buffer solution (PBS) was prepared from monobasic monohydrate sodium phosphate and dibasic anhydrous potassium phosphate (BDH Laboratory Supplies, Poole, England).

Fresh 0.4 M NEM stock solution was prepared in MilliQ water daily. 0.4 M serine/borate complex (SBC) stock solution was prepared in 1 M PBS pH 8.0. GGT was prepared in 0.1 M PBS pH 8.0. 0.2 M GG stock solution was prepared in 1 M PBS pH 8.0. CuSO_4 was prepared in MilliQ water.

Stock solution of GSNO (N4148, CAS No. 57564-91-7, Sigma) was prepared in 0.1 M PBS pH 8.0, 0.5 mM DTPA. As such, GSNO standard solution was stable during the working day (9 h time tested at room temperature) and for 4 weeks if stored at -80°C .

S-Nitrosocysteinylglycine (GCNO), S-nitrosocysteine (CysNO) and S-nitrosohomocysteine (HCysNO) were prepared by reacting 1 M NaNO_2 in H_2O with 1.1 M thiols in 0.5 M HCl, 0.5 mM EDTA at 0°C for 40 min [15]. RSNOs were diluted in 0.1 M PBS pH 8.0, 0.5 mM DTPA and stability was tested before any other use.

Methanol for liquid chromatographic analysis was purchased from Carlo Erba (Rodano, MI, Italy). Water deionized with a Milli-Q system (Millipore, Bedford, MA, USA) was used throughout.

Spectrophotometric analysis

Concentrations of GSNO, CysNO, HCysNO and GCNO in 0.1 M PBS, pH 8.0, at $21 \pm 0.1^\circ\text{C}$ were calculated from the absorbance at 334 nm [16] by a Beckman DU-600 spectrophotometer.

Fluorimetric analysis

A fluorescence spectrophotometer (Perkin-Elmer LS-45 Luminescence Spectrophotometer) was used for method optimization. Slit width was 10 nm for both excitation ($\lambda_{\text{ex}} = 485$ nm) and emission ($\lambda_{\text{em}} = 515$ nm). Measurements were performed in 1 mL, 1 cm path length cuvette. The concentration of DAF-2 standard solution used for determinations was optimized between 0.025 and 0.5 μM . The commercial product DAF-2T gave a calibration curve linear in the range 1–20 nM (slope = 74.4 ± 1.7 , $R = 0.9992$, $N = 5$) and a detection limit of 1 nM.

Measurement of GGT activity

GGT activity was determined by following the hydrolysis of γ -glutamylcarboxy-*p*-nitroanilide (Glu-cp-NA) as previously described [17]. Briefly, purified enzyme was incubated with 1 mM Glu-cp-NA and 20 mM GG in PBS (0.1 M, pH 8.0) at 37°C and the absorbance increase at 405 nm was monitored. Enzyme activity was determined using a molar absorption coefficient of $8800 \text{ M}^{-1}\text{cm}^{-1}$ for the formation of *p*-nitroaniline.

Human blood sampling and storage

Plasma aliquots were derived from routine clinical samples obtained from 13 different in-patients (Inst. of Clinical Physiology–CNR, Pisa). Venous blood was collected by venipuncture using EDTA as anticoagulant. After low-speed centrifugation (1500 g, 10 min) at room temperature, plasma samples were stored at -20°C until analysis. One milliliter of plasma was immediately spiked with 10 μL of 50 mM DTPA stock solution (0.5 mM final concentration of DTPA in plasma) and 30 μL of 0.4 M NEM stock solution (12 mM final concentration of NEM). Then, GGT inhibitor serine/borate complex (SBC; 10 mM final concentration) was added before GSNO spike or after various intervals incubation time. The addition of DTPA/NEM/SBC solution avoided losses of GSNO due to transnitrosation reactions, metal- and enzyme-mediated decomposition (see Results).

Before analysis, plasma samples were diluted 1:1 in 0.1 M PBS pH 7.4, 0.5 mM EDTA, loaded onto the sample reservoir of an Amicon Microcon YM-3 centrifugal filter units (cut-off 3000 Da; Millipore, Bedford, MA, USA) and centrifuged at 11000g for 90 min at 4°C , to remove proteins and high molecular weight compounds. Ultrafiltrates were then injected in the chromatographic system. For the experiments of recovery of spiked RSNO in human whole blood, RSNOs were added after the addition of DTPA/NEM/SBC solution.

Instrumental set-up

The HPLC unit consisted of a pump (P4000, ThermoQuest) equipped with a Rheodyne 7125 injector (Rheodyne, Cotati, CA, USA), a 100 μL poly(etheretherketone) (PEEK tubing, Upchurch, Oak Harbor, WA) injection loop and an HPLC column (Gemini RP C₁₈ 250 \times 4.6 mm, silica particle size 5 μm , equipped with a Guard Cartridge KJ0-4282, Phenomenex). The eluent for the chromatographic analysis was 99% 0.02 M PBS pH 6.0/1% methanol (99:1 v/v), flowing in the optimized conditions at 0.8 mL/min. The flow eluting from HPLC column was monitored between 210 and 650 nm by a UV-visible diode array detector (DAD) equipped with

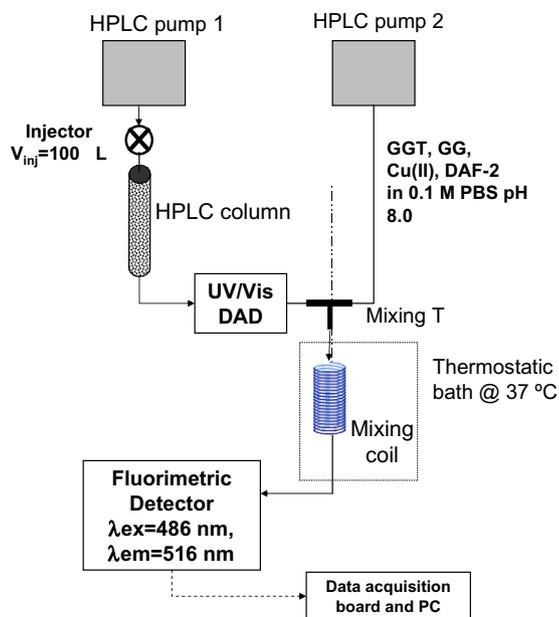


Fig. 1. Schematic diagram of LC-FD system for the determination of GSNO and other RSNOs.

a 5 cm path length flow cell (UV6000, ThermoQuest). The effluent was blended in a low dead-volume mixing-tee with the reaction solution delivered by a second HPLC pump (Jasco PU2080) at a flow rate of 0.3 mL/min, containing 140 mU/mL GGT, 0.03 mM copper sulphate, 1.2 mM GG and 0.37 μ M DAF-2 in 0.1 M PBS pH 8.0 (optimized conditions, concentrations in the reaction coil). The enzymatic reaction proceeded in the optimized conditions in a PFA reaction coil of 2.8 ml, corresponding to a $t_{\text{reaction}} = 2.55$ min, kept at the constant temperature of 37 °C in a water bath (Fig. 1). Thus, the DAF-2T signal was specifically detected by a fluorescence detector (FL3000, ThermoFinnigan) operating at excitation wavelength $\lambda_{\text{ex}} = 486$ nm and emission wavelength $\lambda_{\text{em}} = 516$ nm. Fig. 1 shows a detailed schematic of the LC-DAD-FD system equipped with the post-column reaction system described.

Results

UV spectrophotometric experiments

UV spectrophotometry was used to roughly optimize GG, GGT and CuSO_4 concentrations, prior to optimizing on line reaction conditions (see below). Optimization was performed by a Box-Wilson central composite design, varying GG in the range 5–20 mM, GGT in the range 300–1200 mU/mL, and CuSO_4 between 50 and 250 μ M. Experiments were run in 1 mL cuvettes, in 0.1 M PBS pH 8.0. GG, copper and GGT were added at the established concentrations in the order reported above. Thus, the blank was read in the spectrophotometer, and GSNO was added as last component before starting kinetics data acquisition. By varying the concentration of GGT and GG, and by evaluating the time-dependent plots of the absorbance at 334 nm, we found that 525 mU/mL GGT and 4.5 mM GG (final concentrations) were the optimal conditions for the quantitative decomposition ($\geq 95\%$) of GSNO in about 300 s at 37 °C. In the absence of copper and chelating agents, transition metals present as contaminants in the reaction medium decomposed GCNO slowly. Increasing concentrations of copper rapidly decomposed GCNO. When the reaction medium also included EDTA, GCNO decomposition was not prevented, provided that copper was added in excess with respect to EDTA. A concentration of

100 μ M Cu(II) was sufficient for the quantitative decomposition of 100 μ M GSNO in 300 s, in the presence of 525 mU/mL GGT and 4.5 mM GG. The same experiments performed in the absence of GGT showed that copper by itself (up to 1.2 mM) did not decompose GSNO in the 300 s observation time (*data not shown*).

In the operating conditions employed (4.5 mM GG, 0.1 mM CuSO_4 , 525 mU/mL GGT), the stability of three other S-nitrosothiols was also verified, GCNO, HcysNO, and CysNO. In the absence of GGT GSNO was stable, GCNO was instantaneously decomposed, as previously reported [9], CysNO was instantaneously and fully decomposed as well, and HCysNO was decomposed by about 75% (*data not shown*).

Fluorimetric experiments

GGT-catalysed/copper-mediated GSNO hydrolysis was then investigated in cuvette by fluorescence spectrometry, in order to optimize the DAF-2-mediated detection of the product NO. Experiments were run in 1 mL cuvette in 0.1 M PBS pH 8.0. GG, copper sulphate, GGT and DAF-2 were added at the established concentration. Thus, blank was obtained ($\lambda_{\text{ex}} = 485$ nm, $\lambda_{\text{em}} = 515$ nm) and GSNO was then added as last component, before starting the kinetics data acquisition.

The GGT-catalysed/copper-mediated decomposition of 0.3 μ mol/L GSNO obtained with five different concentrations of DAF-2 (0.5, 0.3, 0.1, 0.05 and 0.025 μ M DAF-2) showed an increase of fluorescence signal as a function of time, which corresponded to formation of DAF-2T following the release of NO from GCNO (*data not shown*). The reaction was complete for a reaction time $t \geq 5$ min for all the concentrations of DAF-2 considered, as expected on the basis of spectrophotometric kinetics at 334 nm. At increasing concentrations of DAF-2 the plateau value F_{10} (=the fluorescence signal after 10 min reaction time) increased as well, indicating an increase of NO detection sensitivity obtained at higher DAF-2 concentrations. However, the overlapping of curves obtained with 0.5 and 0.3 μ M DAF-2 clearly indicated that above 0.3 μ M DAF-2 the rate of reaction (3) no longer depends on DAF-2 concentration, but likely on the rate of formation of the intermediate N_2O_3 [10,11]. At higher DAF-2 concentrations (e.g. 0.5 μ M) the blank signal obtained in the absence of GSNO increased as well, thus affecting the baseline noise signal. Thus, the sensitivity factor (i.e. the slope of the calibration curve) and the baseline noise, which is correlated with the concentration detection limit (LODc), depend on the DAF-2 concentration chosen for the analysis.

Table 1 summarizes the results of data fitting of calibration curves of GSNO standard solutions performed in cuvette in 0.1 mol L⁻¹ PBS pH 8.0, using three different concentrations of DAF-2 (0.05, 0.1 and 0.5 μ M). The LODc of the method in cuvette was 5 nM, and the dynamic linear range was 15–200 nM. It was verified that in the absence of GGT the fluorescence signal did not increase.

Optimization of the reaction conditions for the determination of GSNO and GCNO in plasma

Although GGT specificity makes the proposed approach applicable to in-batch measurements of GSNO, the presence in complex biological matrices (e.g. plasma) of other RSNOs also susceptible to copper-mediated decomposition would require a double determination – with/without GGT – in order to determine (by subtraction) the specific GSNO signal. On the other hand, the set up of a chromatographic separation – coupled to an on line post-column reaction system including reactions (1)–(3) – would make it possible to speciate GSNO as well as other RSNOs susceptible to copper-mediated decomposition. Furthermore, by choosing suitable separation conditions the interference by compounds known

Table 1

Fitting parameters of the linear part of GSNO calibration curves obtained at three different DAF-2 concentrations (50, 100 and 500 nM).

[DAF-2] (nM)	Slope(Volts nM ⁻¹)	Intercept (Volts)	N	R	DLR ^a (nM)
50	1.41 ± 0.09	10.0 ± 5.7	7	0.9966	15–200
100	2.66 ± 0.09	15.1 ± 2.2	8	0.9948	15–200
500	4.87 ± 0.05	66.7 ± 2.5	7	0.9961	15–200

^a Dynamic linear range.

to affect the formation of DAF-2T, such as uric, ascorbic and dehydroascorbic acids [12–14], would be controlled.

Thus, the HPLC separation/post-column reaction procedure was adapted and optimized for GSNO analysis in complex biological matrices. Firstly, the HPLC apparatus coupled to the post-column reaction system and fluorimetric detector (FD) was operated in flow injection mode (=without HPLC column). A solution of commercial GSNO was injected in order to optimize detection conditions: concentration of GGT acceptor GG (ranging 1–20 mmol/L), Cu(II) (100–800 μmol/L), DAF-2 (0.1–2 μmol/L), GGT (0.2–1.5 U/mL), as well as temperature of the thermostatic bath.

Subsequently, HPLC/post-column reaction system/FD in the normal configuration mode (=with the HPLC column inserted between the injection valve and the DAD, Fig. 1), was used to optimize the flow rate of HPLC eluent and post-column reaction solution, and the length of the PFA reaction coil. Parameters were chosen in order to maximize the GSNO signal and minimize the peak width at half height ($W_{1/2}$). This was obtained (i) by limiting the post-column dilution of reactants, and (ii) by increasing the post-column reaction time. A 2.7:1 ratio between HPLC eluent flow and post column solution flow gave optimal results. A flow of 0.8 mL/min was suitable for the separation of GSNO and the other RSNOs (see below). A flow of 0.3 mL/min was chosen for the post-column reaction. Fig. 2A shows the LC-FD chromatograms obtained by injecting a fixed concentration of GSNO without post-column reaction coil or with PFA reaction coils of various lengths, corresponding to increasing reaction times (considering a total flow in the reaction coil of 1.1 mL/min). Fig. 2B shows the normalized slope of GSNO calibration curve (left y axis) and the $W_{1/2}$ values (right y axis) as a function of the reaction time.

The slope values of calibration curves reflected the kinetics obtained in experiments in cuvette. $W_{1/2}$ values were linear as a function of reaction time, as expected. We chose a reaction coil of 2.8 mL, which gave a good sensitivity factor (GSNO was decomposed by GGT by about 65–70%) and a $W_{1/2}$ value compatible with HPLC separation of all RSNOs studied.

Table 2 summarizes values of all post-column reaction parameters, optimized for GSNO determination by LC-DAD-FD. Concentrations shown are those in the reaction coil when considering a 3.7 dilution factor, due to the mixing of HPLC flow (0.8 mL/min) with post-column reaction solution (0.3 mL/min).

As discussed above, Cu(II) can rapidly decompose also CysNO and HCysNO. Thus, in the optimized conditions listed in Table 2, the four RSNOs (GSNO, GCNO, CysNO and HCysNO) could be efficiently determined by the method proposed. Fig. 3 shows the LC-FD chromatogram of a mixture of CysNO, HCysNO, GCNO and GSNO solutions. We verified that sodium nitrite and nitrate solutions injected at mM concentrations gave no fluorescence signal (not shown).

Further determinations were performed in presence or absence of GGT in order to verify the selectivity of GGT reaction as to GSNO. In the absence of GGT, GSNO gave a peak about 1/4 of the one obtained in the presence of GGT (data not shown), indicating some degree of not GGT-related GSNO decomposition. This effect may be produced by Cu(II) being simultaneously present in the reaction medium with GG (required as acceptor substrate for GGT) and

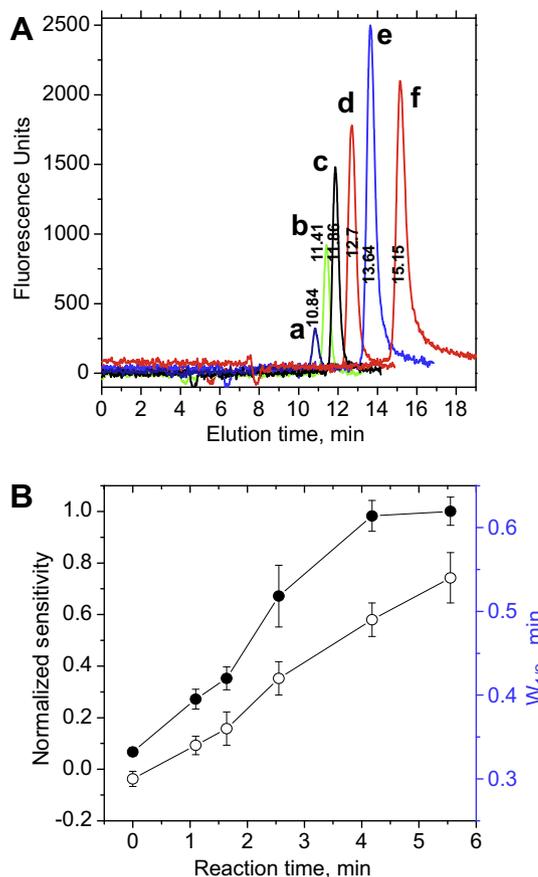


Fig. 2. Optimization of GGT/Cu⁺⁺/DAF-2-mediated GSNO determination. (A) LC-FD chromatograms obtained by injecting a 330 nM GSNO with no post-column reaction coil or PFA reaction coils of various length: (a) no reaction coil, (b) 1.2 mL, (c) 1.8 mL, (d) 2.8 mL, (e) 4.6 mL, (f) 6.1 mL. Operating conditions: 1.2 mM GG, 0.37 μM DAF-2, 140 mU/mL GGT, 0.03 mM Cu(II), HPLC flow 0.8 mL/min, post-column reactant 0.3 mL/min (concentrations in the reaction coil). (B) Normalized slope of GSNO calibration curve (left y axis) and $W_{1/2}$ values (right y axis) as a function of the post-column reaction time.

DAF-2. It is known in fact that formation of adducts with various ligands, including amino acids [18] and DAF-2 itself [16,17], can alter the redox potential of transition metals, often making them more reactive.

Recovery of GSNO from plasma

Fig. 4 shows typical LC-FD and UV absorbance chromatograms of plasma ultrafiltrates. Spikes of RSNOs were added to blood after

Table 2

Optimized values of all parameters for the determination of GSNO and RSNO by HPLC coupled to the post-column reaction system and fluorometric detector.

Parameter	Optimized value
GG concentration	1.2–4.5 mM ^a
Cu(II) concentration	0.03–0.250 mM
DAF-2 concentration	0.37 μM
GGT concentration	140 mU/mL
HPLC flow	0.8 mL/min
Post-column reaction solution flow	0.3 mL/min
PFA reaction coil length	2.8 ml
Temperature of the thermostatic bath	37–41 °C

^a We indicated a range for several parameters that can be varied without affecting significantly the S/N ratio. The sensitivity factor has to be calculated in the chosen reaction conditions.

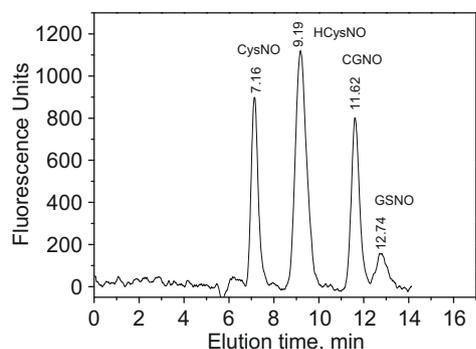


Fig. 3. Determination of different RSNOs by the GGT/Cu⁺⁺/DAF-2 method. LC-FD chromatograms of CysNO, HCysNO, GCNO and GSNO obtained by using the 2.8 mL reaction coil: 115 nM CysNO, 350 nM HCysNO, 80 nM GCNO and 30 nM GSNO.

addition of the stabilizing NEM/DTPA/SBC solution, required to avoid losses due to transnitrosation reactions and metal- and/or GGT-mediated GSNO decomposition.

Table 3 summarizes the results of fitting of RSNOs calibration curves obtained in buffer and analyte addition curve in plasma, in the operating conditions of Table 2, in the 5–1500 nM concentration range. Recovery experiments of RSNOs spiked in plasma evidenced a good recovery and linear analyte addition curves for GSNO (98.6%) and GCNO (99.5%) in the 5–1500 nM range. The limit of quantitation (LOQc) of GSNO and GCNO of the proposed method was 5 nM, with a precision (CV%) of 1.4–5.7% for GSNO and 1–5.8% for GCNO within the dynamic linear range (5–1500 nM) at 0.37 μ M DAF-2 concentration in the post-column reaction coil. LOQc was calculated on the basis of the signal to noise ratio of baseline noise and the slope of calibration curves, and determined experimentally by analysing GSNO and GCNO.

Recoveries of CysNO and HCysNO were instead significantly lower (40–60%) and analyte addition curves were exponential, showing lower recoveries at lower concentrations (5–50 nM) and higher recoveries at higher concentrations (50–1500 nM). Furthermore, the LOQc value determined experimentally for both CysNO and HCysNO was 20 nM. Such low recoveries of CysNO and HCysNO may be due to interference with NO/DAF-2 reaction by ascorbic, dehydroascorbic and uric acids [12–14], which eluted simultaneously to CysNO (ascorbic and dehydroascorbic acid) and HCysNO (uric acid) (Fig. 4), taking into account the shift due to post-column reaction time.

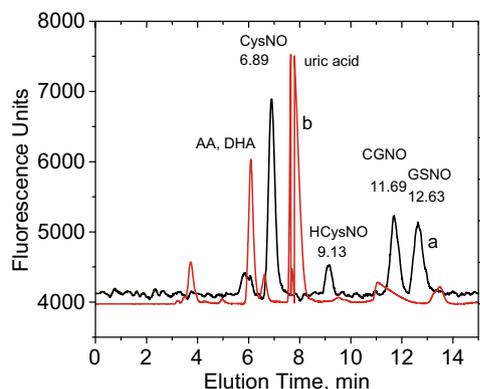


Fig. 4. Recovery of different RSNOs exogenously added to plasma. LC-DAD-FD chromatograms (trace a) and UV absorbance chromatogram at 254 nm (trace b) of a plasma ultrafiltrate sample obtained from blood spiked with 600 nM CysNO, 150 nM HCysNO, 100 nM GCNO and 180 nM GSNO. RSNO concentration found was 310 nM CysNO (recovery 51%), 70 nM HCysNO (recovery 47%), 106 nM GCNO (recovery 106%) and 170 nM GSNO (recovery 94%).

Table 3

Fitting parameters of RSNO calibration curves in 0.1 M PBS pH 8, 0.5 mM DTPA and analyte addition curves in plasma in the optimized operating conditions of Table 2.

RSNO	t_R (min)	Slope (f.u. min nM ⁻¹)	R^2	N
<i>Buffer</i>				
GSNO	12.74	2.77 \pm 0.002	0.9999	10
CNOG	11.62	4.16 \pm 0.005	0.9999	10
CysNO	7.16	2.80 \pm 0.002	0.9999	10
HCysNO	9.19	1.91 \pm 0.005	0.9999	10
<i>Plasma</i>				
GSNO	12.63	2.73 \pm 0.011	0.9999	9
CNOG	11.69	4.16 \pm 0.005	0.9999	7
CysNO	6.89	Not linear	–	–
HCysNO	8.13	Not linear	–	–

Recovery experiments of RSNOs spiked in plasma (approx. 100 nM final conc.) showed that GSNO and GCNO were stable in plasma stored at -80°C up to 3 days (recoveries $\geq 85\%$), while after 1 week the recovery was $\approx 50\%$. Spiked CysNO and HCysNO were unstable: after 24 h the recovery was less than 50%. In plasma ultrafiltrate RSNOs were stable only during the working day. Thus, while GSNO and GCNO can be accurately determined in plasma ultrafiltrate by the LC-FD method, separation and detection of CysNO and HCysNO is not possible in plasma, though potentially feasible in simpler matrices.

Hydrolysis of GSNO by endogenous plasmatic GGT

The good separation of GCNO and GSNO, their good recoveries and determination limit in plasma allowed us to correlate the stability of spiked GSNO with the endogenous content of GGT. In all blood samples used, DTPA and NEM were added before GSNO spike. The metal chelator DTPA should avoid RSNOs losses due to metal-catalysed decomposition [9]. Alkylation by NEM is able to block the reactivity of $-\text{SH}$ groups in about 20 s at neutral pH at room temperature [19]. The addition of NEM to plasma samples should therefore prevent artifactual formation of RSNOs due to transnitrosation reactions from proteins (basically nitroso-albumin) [20] and thiols (in particular GSH) present at micromolar concentration levels in plasma [21]. Moreover, if not blocked by NEM, reduced thiols could reduce adventitious metals present in biological samples: in particular, Cu(I) originating from reduction of Cu(II) could catalyse the decomposition of GSNO [7]. Fig. 5 shows an example of LC-FD chromatogram of human plasma spiked with 60 nM GSNO, and analysed after allowing increasing times before GGT inhibition (=addition of SBC: at 0, 1, 5 and 10 min). $T=0$ was evaluated by adding SBC before the GSNO spike.

As can be seen, when increasing times were allowed before addition of SBC to plasma samples, proportionally lower GSNO recoveries were obtained, while amounts of detectable GCNO correspondingly increased.

From the areas of chromatographic peaks of GCNO ($t_R = 11.90$ min) and GSNO ($t_R = 12.73$ min) and their respective calibration curves, in each sample it was possible to determine at different times the concentrations of residual GSNO and GCNO originated by GGT activity (reaction (1)). Data indicate that depending on the addition time of GGT inhibitor SBC, recoverable GSNO progressively decreased, along with a corresponding increase of GCNO. Fig. 6 shows data obtained with four representative plasma samples (out of 13 examined), characterized by different GGT activities. RSNO concentration was normalized with respect to the GSNO concentration added (60 nM). At $t=0$ the recovery of GSNO was quantitative ($97 \pm 5\%$), while at later times GSNO was converted into GCNO more and more quickly, in correlation with increasing plasma GGT levels.

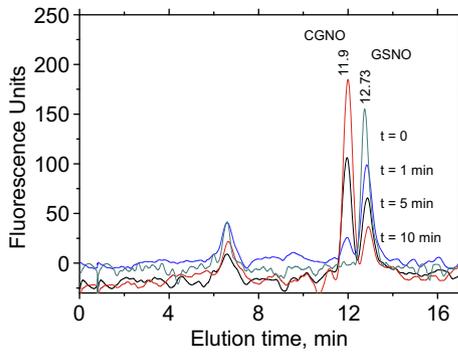


Fig. 5. Interference of endogenous GGT with GSNO determination (I). LC-FD chromatograms of human plasma stabilized by NEM/DTPA, spiked with 60 nM GSNO, and analysed after allowing increasing times before GGT inhibition (=addition of SBC: at 0, 1, 5 and 10 min). $T = 0$ was evaluated by adding the SBC before the GSNO spike. The sample shown contained 114 U/L total GGT. GSNO recovery at $t = 0$ was here 93% taking into account the 1:1 dilution factor of plasma before ultracentrifugation.

Fig. 7 shows the percentage of residual GSNO found in 13 plasma samples after 5 and 10 min with respect to the total GSNO spiked, as a function of plasma GGT activity.

An inverse correlation was observed between plasma GGT activities and percentages of recovered GSNO (linear correlation parameters after 5 min: intercept $87 \pm 4\%$; slope $= -0.18 \pm 0.02$;

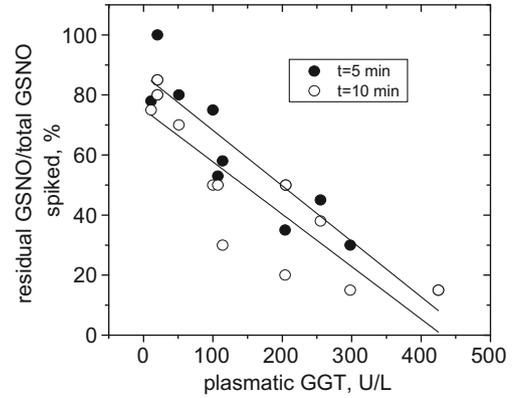


Fig. 7. Interference of endogenous GGT with GSNO determination (III). Recovery of GSNO after 5 (closed circles) and 10 min GGT inhibition time (open circles), as a function of basal GGT activities present in plasma. Recovery was reported as percentage of the concentration of GSNO found over the total amount spiked. Linear correlation parameters after 5 min: intercept $87 \pm 4\%$; slope $= -0.18 \pm 0.02$; $R^2 = 0.8696$; linear correlation after 10 min: intercept $75 \pm 6\%$; slope $= -0.17 \pm 0.03$; $R^2 = 0.7410$.

$R^2 = 0.8696$). A positive correlation was found, instead, between plasma GGT activity and the formation of GCNO (linear correlation parameters after 10 min: slope $= 0.23 \pm 0.03$; $R^2 = 0.8021$) (data not shown). The concentration ($[GSNO] + [GCNO]$) over 10 min was not

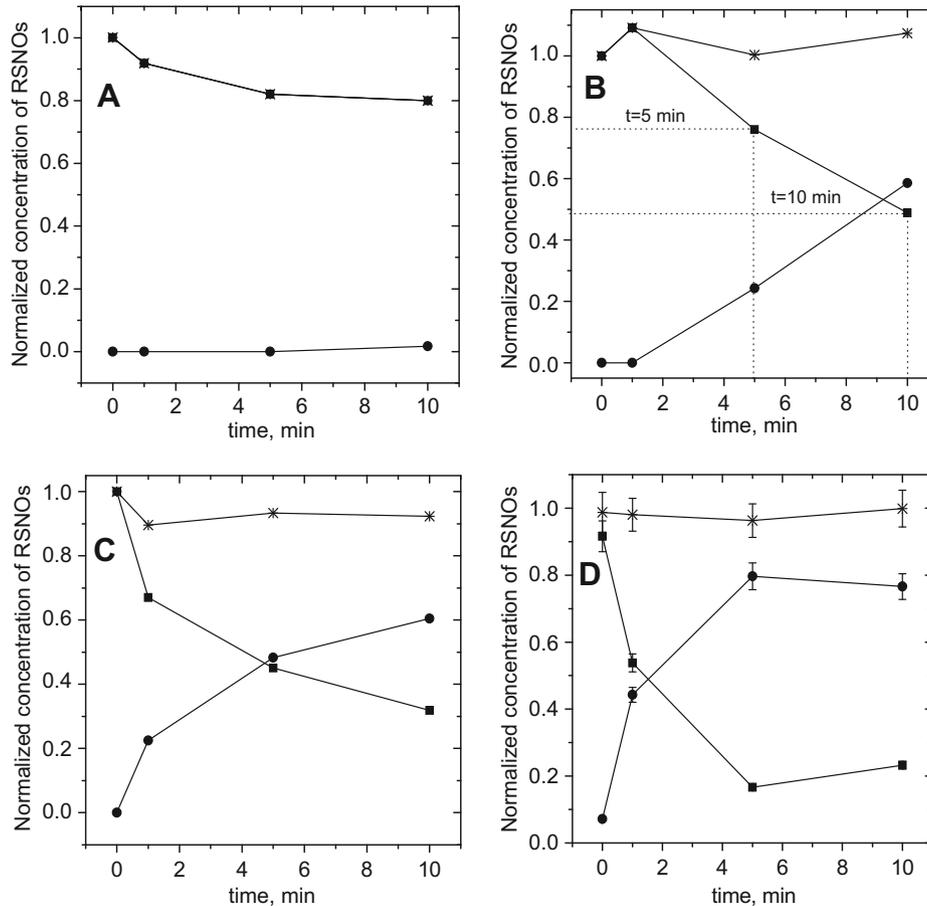


Fig. 6. Interference of endogenous GGT with GSNO determination (II). GSNO was exogenously added and RSNOs were determined in four representative plasma samples characterized by different GGT activities. Data are reported as a function of GGT inhibition times by SBC. Plasma GGT activities: 20 U/L (A), 100 U/L (B), 255 U/L (C) and 425 U/L (D). RSN concentrations were normalized with respect to the GSNO concentration added (100 nM final conc.). Squares: GSNO, circles: GCNO, triangles: GSNO + GCNO. Representative error bars (SD) are reported in panel D ($N = 3$ replicates).

significantly different from the concentration of spiked GSNO, thus indicating a quantitative conversion of GSNO into GCNO.

Discussion

The results of the present study highlight a fundamental aspect to be considered in GSNO research, i.e. the patho-physiological role of endogenous GGT activity possibly (and variably) present in samples. Data reported in a previous paper from our laboratory, concerned with the kinetics of GGT with respect to GSNO as a substrate and glycylglycine as acceptor co-substrate [8], suggested that plasmatic GGT can be a factor in catabolism of GSNO in blood, where numerous endogenous, physiological acceptor co-substrates are available. Altogether our observations are of high relevance as far as GSNO determination: if – besides the inhibition of transnitrosation reactions and metal-induced decomposition – endogenous GGT activity is not efficiently inhibited during sampling (pre-analytical step), the analyte may be lost during the following 1–10 min, directly depending on the levels of GGT present in plasma. This observation may explain the elusiveness of GSNO determination reported in the literature, despite the significant methodological advances achieved over the recent years.

Thus, SBC at the indicated concentration of 10 mM should be used during blood sampling in order to avoid GGT-related GSNO loss, as previously envisaged [22]. Immediate and efficient GGT inhibition is however a more complex issue than apparent. SBC aliquots should perhaps be prepared in *ad hoc* sampling vessels, possibly of the vacuum-type. Using the present method or another recently published by us [22], plasma GSNO concentrations were found to range in between 30 and 300 nM; these data are however to be regarded as preliminary, and subject to confirmation. Moreover, a further aspect liable to affect determination is given by possible haemolysis: even minimal levels could in fact contribute unpredictable amounts of erythrocytic GSNO to the measured values. Studies are in progress in our laboratory in order to investigate these points comprehensively.

In conclusion, a liquid chromatographic system coupled with a post column injection system delivering a buffer solution containing GGT, GG, copper and DAF-2 allowed us to fluorimetrically determine and speciate GSNO and other RSNOs in plasma ultrafiltrate. The idea of using GGT as a reagent to promote the on line hydrolysis of GSNO into GCNO, which in turn is labile to copper-mediated decomposition, represents a novel approach to the long debated analytical problem of specific GSNO determination in biological samples. GSNO and GCNO were determined with a LOQc of 5 nM, a precision (CV) of 1–6% in the 5–1500 nM dynamic linear range, and a recovery 98.6% (GSNO) and 99.5% (GCNO) in plasma ultrafiltrate. The method allowed the appreciation of a critical aspect never investigated before, i.e. the direct interference of (endogenous) GGT activity with detectable GSNO levels. Last but not least, the method has the advantage of using reagents by far

less toxic or environmentally hazardous as compared to HgCl₂ salts, generally used in current procedures for RSNO decomposition in the 0.1–0.2 mM concentration range.

Conflict of interest

Authors C.V., Aldo P., R.B., M.F., A.P. and E.B. have patented the described procedures (Patent No. PCT/IB2007/002090, July 2007). Authors Aldo P., M.F., A.P. and E.B. are members in a University Spin-off company (Sorta Srl, www.sorta-biomedical.com) devoted to contract research based on different analytical services, including the one presented here.

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