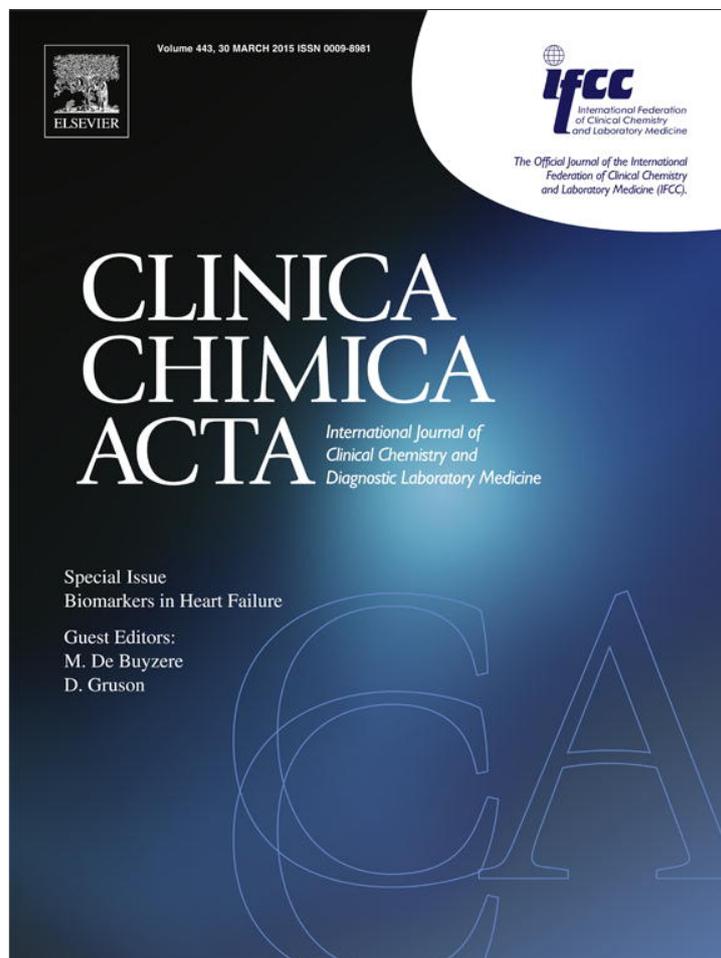


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Invited critical review

Cardiac biomarker testing in the clinical laboratory: Where do we stand? General overview of the methodology with special emphasis on natriuretic peptides



Aldo Clerico*, Claudio Passino, Maria Franzini, Michele Emdin

Scuola Superiore Sant'Anna and Fondazione G. Monasterio CNR – Regione Toscana, Pisa, Italy

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ABSTRACT

Diagnosis of heart failure (HF) is not based on a single test, but on a combination of history, physical examination and appropriate investigations. For these reasons, the accuracy of diagnosis by clinical means alone is often inadequate, especially in the early, asymptomatic stages of the HF. Thus, there is an increasing interest in the development of new cardiovascular biomarkers and, consequently, a great number of laboratory tests have recently been proposed for their assay. The aim of this article is to provide a general overview on the biomarkers, recommended by international guidelines, for the diagnosis, risk stratification, and follow-up of patients with HF. Cardiac natriuretic peptides and in particular the B-type related peptides, which are considered to be the first line biomarker for HF by international guidelines, will be discussed with special emphasis.

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1. Introduction

Estimates on the prevalence of symptomatic heart failure (HF) in the general European and North American population range from 0.4% to 2% [1–5]; with age, HF incidence and prevalence increase steeply, approaching 1 in 1000 among people over the age of 65 [1–5]. From an economic point of view, compared to other diagnoses and treatments, HF is the primary expenditure in Medicare in the US [4], and in healthcare setting across European countries [1–3].

Despite the remarkable advances made during the past 50 years in understanding and treating the disease [6,7], HF continues to have a poor prognosis: approximately up to 40% of patients diagnosed with severe heart failure (NYHA class III–IV or ACC/AHA stage D) in the European and North American population die within one year, with survival rates similar to those of colon cancer, and worse than those of breast or prostate cancer [1–5].

About 20 years ago, Braunwald and Bristow [8] suggested the intriguing hypothesis that it may be possible to reverse the process of HF, which had long been considered to be irreversible and amenable only to palliative therapy. According to this hypothesis, the intrinsic defects in myocardial contraction featured by some patients with chronic HF could be partially reversed by connecting the patient to a ventricular

* Corresponding author at: Dept. of Laboratory Medicine, Fondazione G. Monasterio CNR – Regione Toscana, Via Giuseppe Moruzzi 1, 56124 Pisa, Italy. Tel.: +39 0585 493569; fax: +39 0585 493652.

assist device for several months [9] and/or using an appropriate pharmacological treatment [8]. In particular, it is now well documented that patients with chronic HF, treated with β -adrenergic blocking agents, added to background therapy with ACE inhibitors, improve the systolic function and may reverse cardiac remodeling, leading a better clinical outcomes, including prolonged survival and reduced hospitalizations [1–5]. Thus, the view of chronic HF as an irreversible, end-stage process is being replaced by the concept that intrinsic defects of function and structure afflicting the chronically failing heart can be addressed through appropriate therapy [6]. From a theoretical point of view, we can indeed assume that it is easier to arrest – or even reverse – a progressive process such as HF if action is taken in the earliest phase of the cardiac alteration.

In order to emphasize both the development and progression of the disease, the ACC/AHA guidelines for the diagnosis and management of chronic HF in the adult recommend a classification of HF based on 4 stages from A to D (Fig. 1) [4]. The first two stages (A and B) do not include symptomatic patients in an attempt to underscore to healthcare providers the importance of an early identification of patients who are at risk for developing HF. In particular, patients in stage A have only risk factors without structural or functional alterations of ventricular myocardial, while those in stage B show cardiac structural (such as hypertrophy) and/or functional (such as impaired left ventricular

dysfunction) alterations. The last two stages C and D identify instead symptomatic patients. Since early identification of individuals and risk stratification and diagnosis can be achieved today through the measurement of specific disease or risk markers, an increasing number of new cardiovascular biomarkers have been proposed, as previously reviewed in detail [9–17].

The aim of this review article is to provide a general outline on the methodology of the biomarkers recommended by international guidelines for the diagnosis, risk stratification, and follow-up of patients with HF, with special emphasis on natriuretic peptides, which are considered to be the most useful biomarker for HF.

2. The clinical relevance of biochemical biomarkers in heart failure

HF is defined as a syndrome, resulting from any structural or functional cardiac disorder that impairs the ability of the heart to function as a pump to support a physiological circulation [1–5]. The diagnosis of HF is not based on one single test [1,2]. Positive history and some physical signs (such as orthopnea, rales, third heart sound or jugular vein distension) share a good diagnostic specificity, but also a poor sensitivity in diagnosing acute congestive HF (Table 1) [16,17]. Therefore, the diagnosis of both acute and chronic HF relies on clinical judgment based on a combination of history, physical examination and

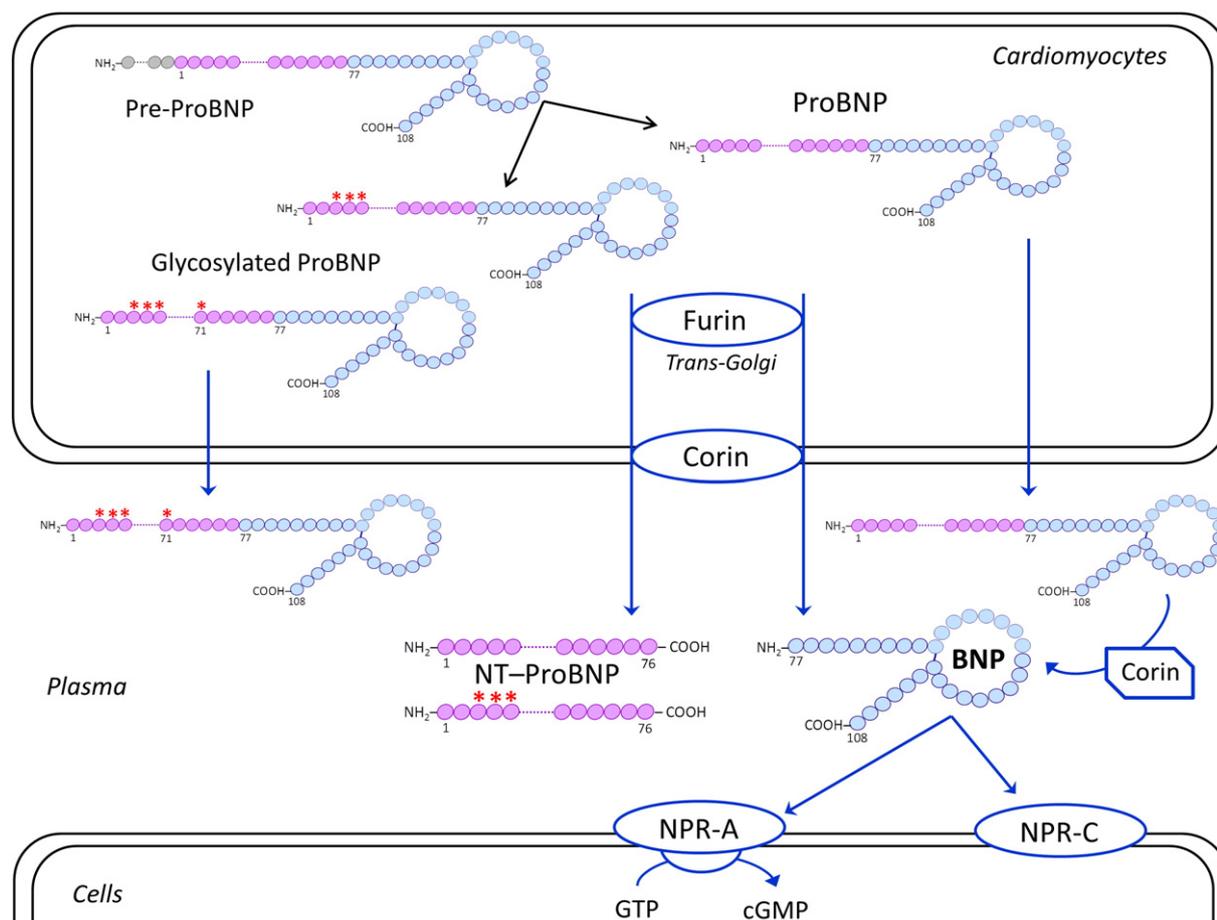


Fig. 1. Schematic representation of biosynthesis, secretion and distribution of B-type related natriuretic peptides. Human BNP is synthesized as a 134-amino acid (aa) precursor protein (pre-proBNP), including a signal peptide of 26 amino acids (grey), and is subsequently processed to form a 108-aa pro-peptide, named proBNP. The proBNP can be enzymatically cleaved by pro-protein convertases produced in the cardiomyocytes, such as corin and furin, mainly located in the trans-Golgi network and on the plasma membrane, respectively [116]. ProBNP is thus processed to form the 76-aa N-terminal peptide (NT-proBNP, violet) and the biologically active 32-aa C-terminal peptide (BNP, light blue), which are both secreted into plasma. Some of the proBNP is O-glycosylated within the Golgi apparatus. Proteolytic cleavage occurs either on or not O-glycosylated proBNP. But, if O-glycans bind to the threonine at position 71 the proBNP will not be processed by furin and corin, thus glycosylated proBNP will be secreted into circulation. Finally, also not glycosylated proBNP can be released as unprocessed peptide. However, the latter can be cleaved into NT-proBNP and BNP by plasmatic corin [117–120]. Only BNP_{1–32}, which is the active hormone, is able to bind the specific receptors, NPR-A and NPR-C. NPR-A is a guanylate cyclase-coupled receptor, which mediates the biological effects of cardiac natriuretic peptides. NPR-C, not coupled to a guanylate cyclase, has essentially a clearance function for all natriuretic peptides.

Table 1

Accuracy of history and physical findings in diagnosing acute congestive HF (modified from references [16,17]).

Variable	Sensitivity	Specificity	Accuracy
History of HF	62	94	80
Dyspnea	56	53	54
Orthopnea	47	88	72
Rales	56	80	70
Third heart sound	20	90	66
Jugular vein distension	39	94	72
Edema	67	68	68

appropriate investigations, as recommended by all international guidelines [1–5,18,19].

According to the most recent international guidelines [18,19], natriuretic peptides, and in particular the peptides related to the B-type cardiac peptide hormone (such as BNP and NT-proBNP), are recommended as the first line biomarker for the diagnosis of both acute and chronic HF. The measurement of BNP and NT-proBNP is useful in supporting clinical judgment for the diagnosis or exclusion of HF in the setting of chronic ambulatory HF or acute decompensated HF (with the maximum degree of class of recommendation I and level of evidence A) [19]. The value of natriuretic peptide testing is particularly significant when the etiology of dyspnea is unclear. Indeed, all international guidelines, starting from the first years of this century, state that lower values of BNP or NT-proBNP actually exclude the presence of HF and higher values have reasonably high positive predictive value to diagnose HF [1–4,18,19].

From a pathophysiological point of view, it is important to underline that the cardiac endocrine function is an essential component of the integrated systems of the body and, thus, plays a pivotal role in fluid, electrolyte and hemodynamic homeostasis [20,21]. A continuous and intense information exchange flows from the endocrine heart system to nervous and immunological systems and to other organs, including the kidney, endocrine glands, liver, adipose tissue, immunocompetent cells, and *vice versa*. This close link between cardiac natriuretic peptide system and counter-regulatory systems could explain the increase in circulating levels of BNP/NT-proBNP in some noncardiac-related clinical conditions [20,21] (Table 2).

As far as the follow-up of HF patients is concerned, BNP and NT-proBNP levels mirror the effectiveness of the treatment of acute or

chronic HF, with lowering of levels over time associated with better clinical outcomes [22,23]. Even if most trials examining the strategy of biomarker “guided” HF management were small and underpowered, at present time, 3 comprehensive meta-analyses concluded that BNP-guided therapy reduces all-cause mortality and cardiovascular hospitalization in patients with HF compared with usual clinical care, especially in patients <75 years of age with co-morbidities [19,21,22,24–27].

As far as the stratification of cardiovascular risk in HF patients is concerned, measurement of natriuretic peptides is recommended by the most recent guidelines with the maximum degree of evidence (class I and level A) also for the prognosis in HF patient [19]. Several studies have demonstrated that increased circulating levels of cardiac troponin I (cTnI) and T (cTnT) – especially using high-sensitivity methods [28,29] – are found in patients with HF, who often do not present obvious myocardial ischemia or underlying coronary artery disease [30–39]. These findings [30–39] suggest that increased cTnI and cTnT in these patients could be caused by cardiomyocyte injury or necrosis. In chronic or acute decompensated HF, elevated cardiac troponin levels are associated with worse clinical outcomes and mortality. Indeed, HF patients, showing a significant and lasting decrease in troponin levels after appropriate pharmacological treatment have a better prognosis compared to those did not show any or only transient decrease [31,38]. Based on these results [30–39], the latest guidelines recommend that troponin I or T be routinely measured, in addition to natriuretic peptides, in patients presenting with acutely decompensated HF for evaluating risk stratification, with the maximum degree of evidence (class I and level A) [19].

In addition to natriuretic peptides and troponins, a huge number of other biomarkers have been suggested for the prognostic value in HF; for example, those related to pro-inflammatory mechanisms, oxidative stress, cachexia, neuro-hormonal dysfunction, and myocardial remodeling, as previously reviewed in detail [9–17]. As an example, a list of some of these suggested biochemical biomarkers is reported in Table 3. In particular, several studies have suggested that biomarkers of myocardial fibrosis, such as galectin-3 [40–53] and soluble ST2 [54–64], are predictive of hospitalization and death in patients with HF. Accordingly [40–64], the most recent guidelines [19] also suggest the use of biomarkers of myocardial fibrosis for additive risk stratification, although with a lower degree of evidence compared to natriuretic peptides and troponins, in both ambulatory (class IIb, level B) and acute (class IIb, level A) HF patients. In Table 4, we summarized

Table 2

Selected causes of elevated natriuretic peptide concentrations (according to references [19–21]).

<i>Cardiac</i>
• Heart failure, including right ventricular syndromes
• Acute coronary syndrome
• Heart muscle disease, including LVH
• Valvular heart disease
• Pericardial disease
• Atrial fibrillation
• Myocarditis
• Cardiac surgery
• Cardioversion
<i>Noncardiac</i>
• Advancing age
• Anemia
• Renal failure
• Liver disease
• Pulmonary: obstructive sleep apnea, severe pneumonia, pulmonary hypertension
• Endocrine diseases (such as hyperthyroidism and primitive or secondary hyperaldosteronism)
• Chronic inflammatory diseases (such as amyloidosis)
• Critical illness
• Bacterial sepsis
• Severe burns
• Toxic-metabolic insults, including cancer chemotherapy and envenomation

Table 3

Suggested prognostic biomarkers for HF.

<i>Neuro-hormones</i>
Natriuretic peptides (ANP, BNP, CNP and related peptides)
Thyroid hormones
Renin–angiotensin–aldosterone system
Catecholamines
Endothelins
Adrenomedullin
Urocortin
Leptin
Adiponectin
<i>Cardiac damage biomarkers</i>
Cardiac troponins, cTnI and cTnT
<i>Remodeling and inflammatory biomarkers</i>
Matrix metalloproteinases (MMPs)
Adhesion molecules (ICAM, VICAM, selectin-P)
C-Reactive Protein (CRP)
Cytokines and related receptor family (IL-2, IL-6, IL-8, TNF- α , ST2)
Pentraxin 3
Galectin 3
<i>Oxidative stress molecular biomarkers</i>
Gamma-glutamyltransferases (GGT)
Reactive oxygen species (ROS), Plasma oxidized LDL
<i>Cachexia biomarkers</i>
Alterations of hypophyseal–suprarenal axis
IGF-1 and GH

Table 4
Pathophysiological and methodological characteristics of biomarkers, which are currently available and recommended by international guidelines [19].

Biomarker type	Pathophysiological relevance in HF	Clinical relevance in HF (Level of evidence)	Analytical methods
<i>ANP and BNP and their related peptides</i>	Cardiac endocrine response to changes in hemodynamics, and to neuro-hormonal and immune systems disturbances	Diagnosis of exclusion of HF (IA) Prognosis of HF (IA) Risk stratification (IA) Follow-up of HF patients (IIA) Additive risk stratification (IA)	Several validated immunoassay methods using automated platforms
<i>Cardiac troponins I and T</i>	Specific marker of injury and damage of cardiomyocytes	Additive risk stratification (IIB)	Several validated immunoassay methods using automated platforms
<i>Galactin –3</i>	Noncardiac specific biomarker related to myocardial remodeling and fibrosis	Additive risk stratification (IIB)	Some immunoassay methods using automated platforms
<i>sST2</i>	Noncardiac specific biomarker related to myocardial remodeling and fibrosis	Additive risk stratification (IIB)	ELISA method

some pathophysiological and methodological characteristics of biomarkers, which are currently available and recommended by the most recent international guidelines for HF according to evidence based medicine principles [19].

3. The multi-markers (MM) approach to cardiovascular risk evaluation

Based on the knowledge acquired so far, it is likely that HF therapy in the future will rely on strategies that combine multiple biomarkers [9,13,16]. This methodological known as the multi-marker (MM) approach or global risk model, is today considered the best model for risk prediction in the individual patient with cardiovascular disease [13,65–67]. Unfortunately, the setup of an adequate MM model is currently complicated by some theoretical and methodological difficulties [68]. According to the MM approach, each biomarker should contribute independently to the diagnostic and prognostic accuracy in a multiple regression model, and ultimately lead to a better outcome for the patient.

In 2010, an expert panel from the American Heart Association established the criteria for the evaluation of novel markers of cardiovascular risk [69]. The panel stated that an adequate evaluation of a novel risk marker should require (i) a sound research design, (ii) a representative at-risk population, and (iii) an adequate number of outcome events. Studies of a novel marker should report the degree to which it adds to the prognostic information provided by standard risk markers. Because no single statistical measure can provide all the information needed to assess a novel marker, studies should also report on measures of both discrimination and accuracy. Furthermore, the clinical value of a marker should be assessed by its effect on patient management and outcomes.

In general, a novel risk marker should be evaluated in several phases according to Evidence-Based Laboratory Medicine principles [70], including initial proof of concept, prospective validation in independent populations, documentation of incremental information when added to standard risk markers, assessment of effects on patient management and outcomes, and ultimately, cost-effectiveness. Biomarkers that do not change the management of a disease are probably unable to significantly affect patient outcome and are thus very seldom cost-effective (judged in terms of quality-adjusted life-years gained) [9,13,71]. Randomized trials are the gold standard for establishing the effectiveness of biomarker-guided strategies [72]. Unfortunately, there are few examples of such trials in cardiology, particularly in the primary prevention setting [72]. Indeed, the lack of well-designed randomized clinical trials explains the relatively low degree of evidence (i.e., class IIa, level B) assigned to the BNP-guided therapy in patients with chronic HF even by the most recent guidelines [19]. However, some pivotal randomized clinical trials on BNP-guided therapy are now in progress [27,73]. The results of these studies will hopefully spread more light on the real usefulness

of this strategy in HF patients, pushing the adoption of BNP-guided therapy in the management of HF patients.

4. Cardiac biomarker testing in the clinical laboratory: where we stand

Cardiovascular biochemical biomarkers are usually measured by means of non-competitive immunometric assay methods, using a combination of two (or more) antisera or monoclonal antibodies specific for separate epitopes of tested biomarkers [9,74]. However, the set-up of a reliable and robust measurement for cardiovascular biomarkers actually presents a very difficult challenger for the experts in laboratory medicine. The desirable characteristics for an “ideal” circulating cardiovascular biomarker, measured by laboratory test, are reported in Table 5.

Cardiovascular biomarkers usually are peptides or proteins (including natriuretic peptides, cardiac troponins, galectin-3 and ST2), which are present in tissues and body fluids at very low concentrations in healthy subjects (i.e., in the range of ng/L). As a result, immunoassay methods with very high analytical sensitivity (i.e., low detection limit, LoD, of about 1 pg/tube or even lower) are necessary to measure with an acceptable analytical imprecision the circulating levels of some cardiac biomarkers (such as troponins and natriuretic peptides) in healthy subjects, especially in pediatric age [28,29,75–78].

Furthermore, some peptide biomarkers (such as BNP) usually share a family of related peptides *in vivo* and *in vitro* [21,79–81], while some protein biomarkers (such as troponins) exhibit considerable chemical and structural heterogeneity in blood of both healthy subjects of HF patients [82–84]. These heterogeneous peptides and proteins can differently cross-react with the antibodies used in immunoassay systems, affecting the accuracy of the measurement. As a result, it is not surprising that there are large systematic differences between the circulating

Table 5
Desirable features of an ideal circulating cardiovascular biomarker measured by a laboratory test.

– Laboratory test acceptable to patient
– Stability <i>in vivo</i> and <i>in vitro</i> of the biomarker
– Adequate analytical sensitivity (functional sensitivity) of laboratory test
– Good degree in reproducibility and accuracy of laboratory
– Easy to perform
– Complete automation of assay
– International standardization of the laboratory test
– Low cost
– Low biological variation of the biomarker
– Cardiac specificity
– Circulating levels of biomarker closely related to the pathophysiological mechanisms of cardiac disease
– Circulating levels of the biomarker closely related to the therapeutic interventions
– Reference range and cut off values tested for gender, age, and ethnicity dependence
– Diagnostic and prognostic accuracy tested by randomized clinical trials
– Cost–benefit ratio favorable tested by randomized clinical trials

levels of biomarkers measured by immunoassay methods for both cTnI [83–85] and BNP [79,86–91].

For example, Wu et al. [92] showed since 1998 that commercial immunoassays generated different results for a given protein concentration due to multiple complex forms of cTnI. According to Apple [93], these data actually demonstrated that it was not possible to compare absolute concentrations obtained with assays from different manufacturers. Commercially available cTnI assays use various standard materials and antibodies with different epitope specificities [83–85]. Hence, troponin I assays may yield results that are unique to a certain method or instrument to the point that values for a same patient sample may differ depending on the assay and platform used [84,93]. Due to the heterogeneity between cTnI methods and without an adequate standardization, reference values and decision limits should be determined separately for each method and not be extrapolated from other assays. In terms of clinical practice, this situation is obviously confusing, especially when patients are referred to different laboratories that use different cTnI methods. Starting 2001 a study group began on the behalf of some international organizations (such as AACC and IFCC) a process for standardization of cTnI immunoassay methods in order to establish a reference measurement procedure and materials [84,95].

5. Standardization or harmonization

Standardization of peptide and protein immunoassays, such as cTnI methods, is a very complicated task [95]. A complete standardization approach needs an accepted reference measurement procedures (RPM) and reference materials for the TnI, which however at present are still not available. Indeed, the term standardization can be used only when comparable results among measurement procedures are based on calibration traceability to SI unit using a RMP [83,84,94–96].

After over 10 years of efforts, some Authors think that cTnI assays is unlikely to become standardized [93]. Fred Apple suggests laboratorians and clinicians not get “bogged down” with cTnI standardization, encouraging to aim efforts toward developing a clear understanding of the clinical and analytical evidence for cTnI immunoassays and “to be happy” that the technological improvements that have led to the precise detection of low cTnI concentrations also will lead to better patient care [93].

Although we completely agree with Apple that the standardization of some critical immunoassay methods truly appears to be a “mission impossible” [97], we do however believe that achieving better harmonization (*i.e.*, a reduction of heterogeneity) among the results provided by different methods is possible. In particular, we hypothesize that a better understanding of biochemical characteristics and the pathophysiological role of a candidate biomarker may promote a harmonization process indicating to manufacturers and laboratorians specific targets (*i.e.*, epitopes) for the set-up of more accurate immunoassay methods. A good example for a possible harmonization process is represented by immunoassay methods for cardiac B-type-related natriuretic peptide system.

6. The B-type cardiac natriuretic peptide system

The human BNP gene encodes for a pre-proBNP molecule of 134 amino acid residue, including a signal peptide of 26 amino acids. BNP is cleaved out of a prohormone molecule of 108 amino acids, the proBNP_{1–108} (proBNP). According to the “classical” scenario of BNP production and secretion from cardiomyocytes, before being secreted from cardiomyocytes into the bloodstream, proBNP is split by some proteolytic enzymes (such as corin and/or furin) into two peptides: the biologically inactive NH₂-terminal peptide fragment proBNP_{1–76} (NT-proBNP), and the COOH-terminal peptide fragment proBNP_{77–108} [21] (Fig. 1). The latter is a peptide of 32 amino acids (BNP_{1–32}) and is usually indicated as BNP. This is the active hormone, that is the only able to bind

to the specific natriuretic peptide receptors (named NPR-A, B, C, respectively) [21].

Some recent studies open a new and more complex scenario regarding the pathophysiological and clinical relevance of circulating B-type natriuretic peptides [22]. In addition to the peptide hormone BNP and the inactive peptide NT-proBNP, a huge numbers of circulating proBNP-derived fragments can be identified by chromatographic procedures in human plasma, including the intact and glycosylated forms of the precursor proBNP [98–116] (Fig. 1). Several studies have also demonstrated that intact or glycosylated forms of proBNP constitute a significant portion of immunoreactive B-type-related peptides circulating in plasma of patients with heart failure [98–116]. According to these findings, it is theoretically conceivable that the active hormone (*i.e.*, BNP) may be produced even *in vivo* from the circulating intact precursor proBNP through enzymatic cleavage by some plasma proteases (such as corin) [117–119]. Indeed, a recent study using an *in vivo* rat model demonstrated that processing of human proBNP to active BNP can actually occur in the circulation [120]. The peripheral processing of circulating proBNP could likely be submitted to regulatory mechanisms, which might be impaired in patients with heart failure, opening new perspectives in the treatment of heart failure [80,121]. Indeed, a novel pharmacological target may be the pharmacodynamic action of drugs inducing and/or modulating the maturation of the prohormone into active hormone (*i.e.*, BNP) [122].

From a methodological and analytical points of view, the large heterogeneity of B-type natriuretic peptides circulating in human blood seems to explain the systematic differences among the results provided by immunoassay methods considered specific to the peptide hormone BNP [87,90,91,123,124]. In particular, a recent study, using standard protocols and quality control materials, demonstrated that the IRMA method (by Shionogi's Diagnostic Division, Japan), the ADVIA method for the Centaur platform (by Siemens Health Care Diagnostics) and ST AIA-PACK method for the AIA platform (by TOSOH Corporation, Tokyo, Japan) measured greatly lower (up to the half) BNP values in comparison with other immunoassays, such as the POCT Triage method (by Alere Diagnostics), the BNP Triage Biosite for Access and UniCell DxI platforms (by Beckman Coulter Diagnostics), the MEIA method for the AxSYM platform and the chemiluminescent microparticle immunoassay for ARCHITECT platform (both by Abbotts Diagnostics) [124]. It is interesting to note that the IRMA method by Shionogi, the ADVIA method for Centaur platform, the ST AIA-PACK method (personal communications from TOSOH EUROPE N.V., Tessenderlo, Belgium) use the same antibodies and standard materials supplied by Shionogi's Diagnostic Division [18].

According to the study by Luckenbill et al. [79], a great part of these systematic differences between the different BNP immunoassay systems should be due to the cross-reaction with the glycosylated or not glycosylated proBNP. Liang et al. [98] demonstrated that proBNP constitutes a substantial portion of immunoreactive BNP measured in plasma of HF patients. More recently, Macheret et al. [108], in study using a specific immunoassay method for proBNP [101], demonstrated that this precursor peptide of BNP was detectable in all subjects studied, and its levels were dependent of gender, age, heart rate, and body mass index. Furthermore, these Authors found that the degree of clinical sensitivity and specificity of proBNP assay for the detection of left ventricular dysfunction was comparable to two commercial assays for BNP and NT-proBNP [108]. The results of this study [108] confirm previous reports [101–107] suggesting that the intact precursor of biologically active BNP (*i.e.*, the proBNP) circulates in plasma of both healthy subjects and HF patients.

7. What B-type-related peptide should we measure and why?

According to this new scenario regarding the circulating levels of B-type cardiac natriuretic peptides, there are at least 3 different peptides that could be measured in human plasma samples: the active peptide

hormone, BNP, the inactive N-terminal fragment, NT-proBNP, or the pro-hormone peptide, proBNP [21,121]. These 3 peptides have different biochemical characteristics and pathophysiological relevance (Table 6).

From an analytical point of view, the inactive peptide NT-proBNP and proBNP, are more stable *in vivo* and *in vitro*, with a longer plasma half-life and a lower intra-individual biological variation, than active peptide BNP (Table 4) [18,21,81]. From a pathophysiological point of view, several studies indicate that the inactive peptides, especially proBNP, show an incremental increase of their circulating levels related to progression of HF greater than the active peptide hormone BNP [102,103,110,115,125]. In particular, two studies [115,125], which identified and quantified the individual cardiac natriuretic peptides by means of mass spectrometry, reported that the real levels of the peptide hormone BNP_{1–32} in patients with severe HF are much lower than the BNP concentrations measured by commercially available immunoassay methods. Furthermore, results of another study using mass spectrometry determination actually provide specific evidence for the absence (*i.e.*, concentration below the analytical sensitivity of the measurement) of circulating active peptide BNP_{1–32} in advanced-stage HF patients [126].

According to the analytical characteristics and clinical results discussed above, we would assume that the inactive peptide NT-proBNP and proBNP would be a better biomarker for the progression of HF than the active hormone BNP. However, at present, all the commercially available immunoassay methods considered specific to active peptide BNP significantly cross-react with proBNP [79]. In agreement with these data [79], a recent study [127] has found a good correlation between the BNP and proBNP values measured with commercially available immunoassay methods considered specific for these two peptides in patients with severe HF. Moreover, all the international guidelines state that the commercially available BNP and NT-proBNP immunoassays usually give clinically comparable results when used for diagnosis, prognosis and follow-up of HF patients [18,21,80,88,89,128].

At present, we must realize that the commercially available immunoassay methods considered specific for the active form of B-type cardiac natriuretic peptides present an obvious paradox. From a pathophysiological point of view, it would be better to measure the active peptide BNP (instead of inactive peptide NT-proBNP and proBNP) when we are interested in evaluating the “true biologically active status” of the cardiac endocrine function [21,22]. However, to date, none of the commercially available methods is able to provide such information, accurately, as these methods are greatly affected by inactive peptides the concentrations of which are higher than active hormone in the blood samples of HF patients. In other words, at present time, all the commercially available immunoassay methods for BNP assay are not completely specific for the active form of the peptide.

In conclusion, BNP immunoassay methods show large systematic differences due to the interferences of some inactive peptides, especially the glycosylated and non-glycosylated forms of the precursor peptide, proBNP. The setup of more specific methods for the active peptide BNP_{1–32} could reduce these systematic differences resulting a better harmonization among results.

Table 6
Biochemical and physiological characteristics of BNP, NT-proBNP and proBNP peptides.

	BNP	NT-proBNP	proBNP
Molecular mass	3462 Da	8457 Da ^a	11900 Da ^a
Amino acids	32	76	108
Biological function	Active hormone	Inactive	Pro-hormone
Half life	15–20 min	>60 min	>60 min
Glycosylation	Not glycosylated	Highly glycosylated <i>in vivo</i>	Highly glycosylated <i>in vivo</i>

^a The molecular mass (MM) of NT-proBNP and proBNP depends to the degree of glycosylation of the peptide; in the Table are reported the MM of not glycosylated peptides.

As a future perspective, a more accurate estimation of both production/secretion of B-type related peptides from cardiomyocytes and overall activity of the cardiac endocrine function could be achieved by the testing plasma samples using simultaneously two methods: one specific for the intact precursor proBNP_{1–108}, and the other for active peptide BNP_{1–32} [80,121]. Information obtained by the contemporaneous measurement of proBNP and BNP with specific assays may likely extend our present understanding of pathophysiological mechanisms linking together disease progression and cardiac endocrine dysfunction [80]. However, the clinical usefulness of these new and more specific methods will have to be accurately evaluated by randomized clinical trials in comparison with NT-proBNP and proBNP methods according to the evidence-based medicine principles [69–72].

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